

MALATTIA LINFOPROLIFERATIVA POST-TRAPIANTO

FAD SINCRONA 4 dicembre 2024



PTLD: strategie di prevenzione e monitoraggio nelle diverse tipologie di trapianto. Immunosoppressione, infezioni concomitanti e profilassi antivirale

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PLTD: differences in incidence and time distribution in allo-HSCT and SOT

HSCT

- 0.1-63%. EBMT study:3.2% (1.2% in MRD->11.2% in MMUD)
- Median time from transplantation: 2-4 months (4% later than 12 months)
- Almost 100% of PTLD cases are associated with EBV
- Generally derives from the donor's B lymphocytes

About 20.000 allo-HSCT per year in Europe.

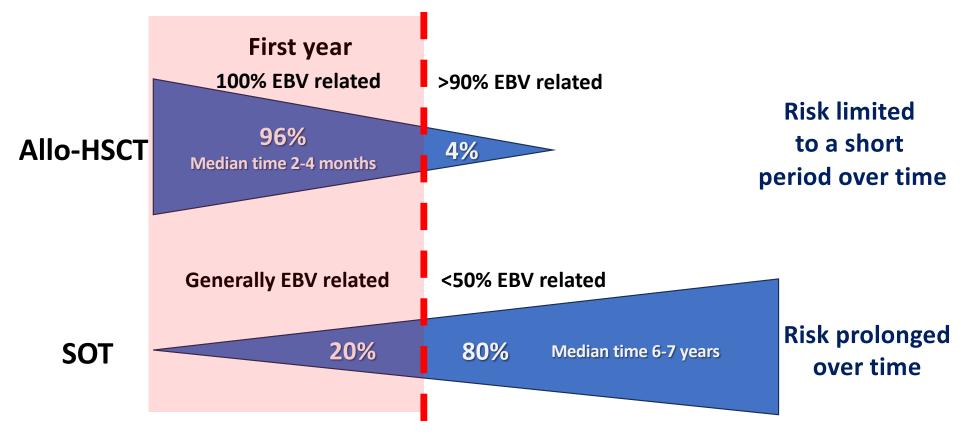
SOT

- 1-33%. Spain study: 1.8% (1.4% in kidney->16.4% in multivisceral transplantation)
- Median time from transplantation: 6.7 y (21% within12 months)
- ~50% of PTLD cases are associated with EBV
- Generally derives from the recipient's B lymphocytes

About 28.000 SOT per year in Europe.

Styczynski, et al. Haematologica 2016;101:803-11 – Jimenez Ubieto et al Blood 2023;142:4490-2 - Tichadou et al Blood 2023; 142:443-5

EBV/PTLD in allo-HSCT and SOT: different epidemiology, different monitoring strategy





Revie

Recent Advances in Adult Post-Transplant Lymphoproliferative Disorder

Mariam Markouli ¹, Fauzia Ullah ², Najiullah Omar ², Anna Apostolopoulou ³, Puneet Dhillon ⁴, Panagiotis Diamantopoulos ¹, Joshua Dower ⁵, Carmelo Gurnari ², Sairah Ahmed ^{6,†} and Danai Dima ^{2,7,*,†}

Table 2. Risk factors for PTLD.

| Post-SOT | Post-alloHSCT |
|--|---|
| Strong Evidence: | Strong Evidence: |
| Intestinal > Lung > Heart > others Multivisceral grafts or graft from deceased donors EBV Seronegative/naive EBV recipient pre-SOT High intensity IST Anti-thymocyte globulin use as part of induction IST | High degree of HLA mismatch HLA-mismatched or unrelated donor Haploidentical donor Umbilical cord blood graft use Type of conditioning regimen T-cell-depleting strategies (in vivo and ex vivo) |
| Weak Evidence: | Anti-thymocyte globulin use |
| a. Non-white ethnicity b. Young recipient and old donor age | Non-myeloablative conditioning regimens 3. Recipient old age > 50 years |
| c. Non-EBV infection | Weak Evidence: |
| d. Recipient HLA-A26 and B38 status | a. Acute GVHD b. History of splenectomy c. Diagnosis of Aplastic Anemia d. Non-EBV infection |

Management of Epstein-Barr Virus infections and post-transplant lymphoproliferative disorders in patients after allogeneic hematopoietic stem cell transplantation: Sixth European Conference on Infections in Leukemia (ECIL-6) guidelines



Jan Styczynski, Walter van der Velden, 2 Christopher P. Fox, 3 Dan Engelhard, 4 Rafael de la Camara, 3 Catherine Cordonnier, 8 and Per Ljungman 7 on behalf of the Sixth European Conference on Infections in Leukemia, a joint venture of the Infectious Diseases Working Party of the European Society of Blood and Marrow Transplantation (EBMT-IDWP), the Infectious Diseases Group of the European Organization for Research and Treatment of Cancer (EORTC-IDG), the International Immunocompromised Host Society (ICHS) and the European Leukemia Net (ELN)

Haematologica 2016 Volume 101(7):803-811

Major risk factors for clinically significant EBV reactivation after allo-HSCT.

- T-cell depletion (in vivo, ex vivo)
- EBV serology
- HLA mismatch
- Severe acute or chronic GvHD

Table 3. Risk factors for EBV-PTLD after HSCT.

Pre-transplant risk factors

- T-cell depletion (either in vivo or ex vivo)
- · EBV serology donor/recipient mismatch
- Cord blood transplantation (CBT)
- · HLA mismatch
- Splenectomy
- Second HSCT

Post-transplant risk factors

- Severe acute (especially steroid-refractory) or chronic GvHD requiring intensive immunosuppressive therapy
- High or rising EBV viral load
- · Treatment with mesenchymal stem cells

Biol Blood Marrow Transplant 25 (2019) 1441-14



Biology of Blood and Marrow Transplantation



journal homepage: www.bbmt.or

Analysis

Risk Factors and Predictive Scoring System For Post-Transplant Lymphoproliferative Disorder after Hematopoietic Stem Cell Transplantation



Ayumi Fujimoto ^{1,2}, Nobuhiro Hiramoto ⁷, Satoshi Yamasaki ³, Yoshihiro Inamoto ⁴, Naoyuki Uchida ⁵, Tetsuo Maeda ⁶, Takehiko Mori ⁷, Yoshinobu Kanda ⁸, Tadakazu Kondo ⁸, Souichi Shiratori ¹⁰, Shigesaburo Miyakoshi ¹¹, Ken Ishiyama ¹², Kazuhiro Ikegame ¹³, Yoshiko Matsuhashi ¹⁴, Junj

Table 2

Characteristics of Allogeneic HSCT

| haracteristics of Allogene | ic HSCT | |
|--------------------------------|-------------------------------|-------------------------|
| Variable | No PTLD Group (N = 39,928) | PTLD Group (N = 267) |
| Conditioning regimen, n (%) | | |
| MAC | 23,680 (60) | 123 (47) |
| RIC | 16,144 (40) | 140 (53) |
| Unknown | 104(0) | 4(0) |
| Donor type, n (%) | | 1.7 |
| MRD | 13,034 (33) | 24 (9) |
| MMRD | 4043 (10) | 52 (19) |
| MURD | 10,135 (25) | 81 (30) |
| MMURD | 1977 (5) | 16 (6) |
| СВ | 10,030 (25) | 88 (33) |
| Unknown | 709(2) | 6(2) |
| Stem cell source, (%) | | |
| BM | 20,063 (50) | 126 (47) |
| PB | 9660 (24) | 52 (19) |
| СВ | 10,030 (25) | 88 (33) |
| Other/unknown | 175 (0)/3 (0) | 1 (0)/0 (0) |
| GVHD prophylaxis, n (%) | | |
| CSP-based | 18,216 (46) | 88 (33) |
| TAC-based | 20,593 (52) | 172 (64) |
| Other | 637 (2) | 4(2) |
| None/unknown | 274 (0)/208 (0) | 0 (0)/3 (1) |
| Use of ATG, n (%) | 3915 (10) | 111 (42) |
| Conditioning only | 3299 (8) | 94 (35) |
| GVHD prophylaxis only | 70 (0) | 1 (0) |
| Acute GVHD treat- ment only | 303 (1) | 6(2) |
| Two or more | 234(1) | 10 (4) |
| No/unknown | 35,870 (90)/152 (0) | 152 (57)/4 (2) |
| Use of alemtuzumab, n (%) | 45/38,895 (<1) | 0/251 (0) |
| Use of ex vivo TCD, n (%) | 291/38,766 (<1) | 5/264 (2) |
| Acute GVHD grade II-IV (%) | | |
| Yes | 13,797 (35) | 115 (43) |
| No/unknown | 22,784 (57)/3347 (8) | 142 (53)/10 (4) |
| Chronic GVHD | | |
| Yes | 12,150 (30) | 88 (33) |
| No/unknown | 18,298 (46)/9480 (24) | 146 (55)/33 (12) |

Table 3Univariate and Multivariate Analyses for the Development of PTLD

| Variable | | Univariate Analysis | | | Multivariate Analysis | | |
|-------------------------------|------|---------------------|-------|------|-----------------------|-------|--|
| | HR | (95% CI) | P | HR | (95% CI) | P | |
| Year of HSCT | | | | | | | |
| 1990-2009 | 1.00 | | | 1.00 | | | |
| 2010-2015 | 2.77 | (2.13-3.61) | <.001 | 1.87 | (1.38-2.52) | <.001 | |
| Disease | | | | | | | |
| AML/MDS | 1.00 | | | 1.00 | | | |
| ALL | .99 | (.69-1.44) | .98 | 1.08 | (.75-1.57) | .68 | |
| CML/MPD | .94 | (.56-1.57) | .81 | 1.55 | (.89- 2.69) | .12 | |
| Lymphoid malignancies | 1,24 | (.88-1.75) | .22 | 1.33 | (.92-1.92) | .13 | |
| AA | 4.95 | (3.47-7.07) | <.001 | 5.19 | (3.32-8.11) | <.001 | |
| Others | 1.91 | (.97-3.76) | .06 | 1.94 | (.97-3.89) | .06 | |
| Conditioning regimen | | | | | | | |
| MAC | 1.00 | | | 1.00 | | | |
| RIC | 2.00 | (1.56-2.55) | <.001 | .82 | (.60-1.12) | .22 | |
| Donor type | | | | | | | |
| MRD | 1.00 | | | 1.00 | | | |
| MMRD | 10.4 | (6.35-17.1) | <.001 | 4.39 | (2.39-8.07) | <.001 | |
| MURD | 4.89 | (3.07-7.79) | <.001 | 4.08 | (2.39-6.99) | <.001 | |
| MMURD | 5.46 | (2.88-10.3) | <.001 | 3.20 | (1.58-6.47) | .001 | |
| СВ | 7.24 | (4.56-11.5) | <.001 | 8.03 | (4.72-13.7) | <.001 | |
| Number of allogeneic HSCT | | | | | | | |
| Two or more | 2.15 | (1.56-2.97) | <.001 | 1.50 | (1.05-2.15) | .03 | |
| GVHD prophylaxis | | | | | | | |
| CSP-based | 1.00 | | | 1.00 | | | |
| TAC-based | 2.07 | (1.59-2.69) | <.001 | .82 | (.59-1.12) | .21 | |
| ATG in a conditioning regimen | | | | | | | |
| Yes | 7.76 | (6.03-9.99) | <.001 | 6.13 | (4.33-8.68) | <.001 | |
| ATG for GVHD treatment* | | | | | | | |
| Yes | 6.87 | (4.00-11.8) | <.001 | 2.09 | (1.17-3.72) | .01 | |
| Acute GVHD grade II-IV* | | | | | | | |
| Yes | 1.83 | (1.43-2.35) | <.001 | 1.93 | (1.48-2.52) | <.001 | |

^{*} ATG for GVHD treatment and acute GVHD grade II-IV were treated as time-dependent variables.

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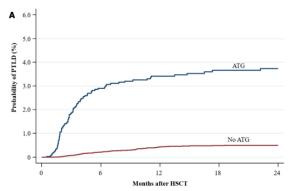
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Risk Factors and Predictive Scoring System For Post-Transplant Lymphoproliferative Disorder after Hematopoietic Stem Cell Transplantation



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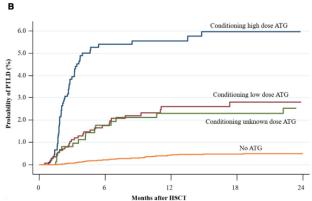


Figure 3. Probability of PTLD with the use of ATG in conditioning. (A) The probability of PTLD was significantly higher in patients who received ATG. (B) Receipt of high-dose ATG (total dose - 2.55 mg/kg Thymoglobulin or -5.0 mg/kg ATG-F) was associated with a significantly higher risk of developing PTLD. By 2 years after HSCT, PTLD developed in 6.050 of patients who received high-dose ATG and in 2.85 of those who received low-dose ATG and in 2.85 of those who received low-dose ATG and in 2.85 of those who received low-dose ATG and in 2.85 of those who received low-dose ATG and in 2.85 of those who received low-dose ATG and in 2.85 of those who received low-dose ATG and in 2.85 of those who received low-dose ATG and in 2.85 of those who received low-dose ATG and in 2.85 of these who received low-dose ATG and in 2.85 of these who received low-dose ATG and in 2.85 of these who received low-dose ATG and in 2.85 of these who received low-dose ATG and in 2.85 of these who received low-dose ATG and in 2.85 of these who received low-dose ATG and in 2.85 of these who received low-dose ATG and in 2.85 of these who received low-dose ATG and in 2.85 of these who received low-dose ATG and in 2.85 of these who received low-dose ATG and in 2.85 of these who received low-dose ATG and in 2.85 of these who received low-dose ATG and in 2.85 of these who received low-dose ATG and in 2.85 of the second low-dose ATG and in 2.85 of the who received low-dose ATG and in 2.85 of the who received low-dose ATG and in 2.85 of the who received low-dose ATG and in 2.85 of the who received low-dose ATG and in 2.85 of the who received low-dose ATG and in 2.85 of the who received low-dose ATG and in 2.85 of the who received low-dose ATG and in 2.85 of the who received low-dose ATG and in 2.85 of the who received low-dose ATG and in 2.85 of the who received low-dose ATG and in 2.85 of the who received low-dose ATG and in 2.85 of the who received low-dose ATG and in 2.85 of the who received low-dose ATG and in 2.85 of the who received low-dose A

PTLD incidence: 0.66%

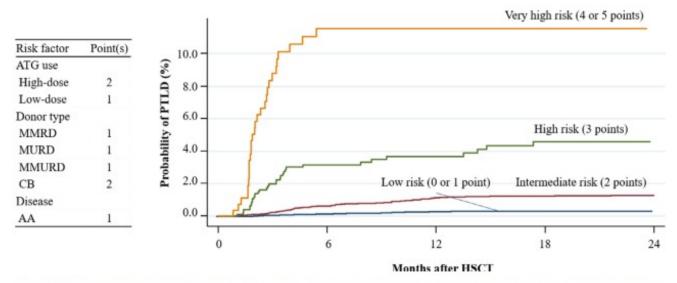


Figure 4. Probability of PTLD by the risk scoring system. Points were assigned for each risk factor as follows: high-dose ATG use, 2 points; low-dose ATG use, 1 point; MMRD, 1 point, MURD, 1 point; MMURD, 1 point; CB, 2 points, and AA, 1 point. The sum of points was used to classify risk groups: 0 or 1 point, low risk; 2 points, intermediate risk; 3 points, high risk; and 4 or 5 points, very high risk. The very-high-risk and high-risk groups had a markedly greater risk of developing PTLD, with probabilities of PTLD at 2 years after HSCT in these risk groups of 11.5% and 4.6%, respectively.

Post-transplant lymphoproliferative disorders, Epstein-Barr virus infection, and disease in solid organ transplantation: Guidelines from the American Society of Transplantation Infectious Diseases Community of Practice

Upton D. Allen^{1,2,3} | Jutta K. Preiksaitis⁴ | on behalf of the AST Infectious Diseases Community of Practice

TABLE 1 Risk Factors for PTLD in solid organ transplant recipients

Early PTLD

Primary EBV infection

Type of organ transplanted (intestine > lung>heart > liver>pancreas > kidney)

Polyclonal anti-lymphocyte antibodies^a

Young recipient age (ie, infants and young children)

Late PTLD

Duration of immunosuppression

Type of organ transplanted

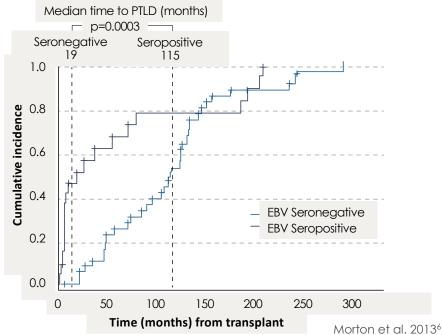
Older recipient age (ie, adults)

- An overwhelming risk factor in most analyses is EBV-seronegativity pre-transplant and primary EBV infection, placing pediatric populations at higher risk of developing PTLD than their adult counterparts
- An increased risk associated with being EBV seronegative in kidney (HR 3.6), and heart (HR 4.0) but found a smaller but significantly increased risk in seronegative liver recipients (HR 1.5).
- Among seronegative pediatric recipients donor seropositivity (D+R-) and donor seronegativity (D-R-) resulted in comparable risks for PTLD at three years post-transplant, perhaps reflecting the high rate of community-acquired infection in children.
- In contrast, in seronegative adult recipients D-Rrecipients trended toward having a lower risk of PTLD than D+R- recipients which received statistical significance when a living donor was used.
- Intestinal transplant recipients appear to have an exceptional high risk of PTLD development, independent of pre-transplant EBV serostatus

EBV status and PTLD development post-SOT

 Pre-transplant EBV seronegativity increases the incidence of PTLD from 10- to 75-fold over that of EBVseropositive recipients^{1,2}

Time to PTLD onset according to EBV status at the time of transplantation³



EBV, Epstein-Barr virus; PTLD, post-transplant lymphoproliferative disorder; SOT, solid organ transplantation.

1. Walker RC, et al. Clin Infect Dis. 1995;20:1346–1353; 2. Cockfield SM. Transpl Infect Dis. 2001;3:70–78; 3. Morton M, et al. Transplantation. 2013;95:470–480.

Management of Epstein-Barr Virus infections and post-transplant lymphoproliferative disorders in patients after allogeneic hematopoietic stem cell transplantation: Sixth European Conference on Infections in Leukemia (ECIL-6) guidelines



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- Prospective monitoring of EBV DNA performed by quantitative PCR is recommended.
- Screening for EBV DNA-emia should start within the first month after allo-HSCT. However, the incidence of EBV-PTLD during the first month after HSCT is estimated to be below 0.2%. Monitoring should continue for at least 4 months after HSCT, with a frequency of at least once a week.
- As the calculated doubling time for EBV might be as short as hours, more frequent sampling in patients with rising EBV DNAemia may be warranted

SPECIAL ISSUE-TRANSPLANT INFECTIOUS DISEASES WILEY Clinical Transplantation. 2019;33:e13652.

Post-transplant lymphoproliferative disorders, Epstein-Barr virus infection, and disease in solid organ transplantation: Guidelines from the American Society of Transplantation Infectious Diseases Community of Practice

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- Studies of the sensitivity and specificity of quantitative EBV viral load for the diagnosis of early PTLD and symptomatic EBV infection are limited
- The use of EBV viral load as a diagnostic test has good sensitivity for detecting EBV-positive early PTLD but misses EBV-negative as well as some cases of localized and donor-derived EBV + PTLD
- However, it had poor specificity, resulting in good negative (greater than 90%) but poor positive predictive value (as low as 28% and not greater than 65%) in these populations
- Elevated and often sustained elevation in EBV loads has been observed in 67%-72% of adult liver, 31%-29% of adult kidney, and 13%-42% (assay dependent) of adult lung transplant recipient EBV-seropositive pre-transplant and appears to be a poor marker of future PTLD risk.

Challenging issues in the monitoring of EBV/PTLD in allo-HSCT and SOT

Allo-HSCT

- Variable risk but in a short time period
- EBV-DNAemi is a sensitive marker of future PTLD
- DNAemia monitoring intensification during the first months from transplant in all transplants
- In the clinical practice EBV-DNAemia monitoring associated to CMV DNAemia monitoring

SOT

- EBV-DNAemia is a good marker of early PTLD in seronegative children and in very high risk SOTs (i.e. intestinal and multivisceral transplant)
- EBV-DNAemia is a poor marker of late PTLD
- Considering the prolonged risk over time, it is difficult to define how prolonged should be the virological monitoring.
- In view of the lack of a good marker for virological monitoring clinical suspicion of late PTLD is crucial



JPIDS 2024:13 (Suppl 1) • S31

A Focused Review of Epstein-Barr Virus Infections and PTLD in Pediatric Transplant Recipients: Guidance From the IPTA and ECIL Guidelines

Masaki Yamada,^{1,1,©} Arnaud G. L'Huillier,^{2,3,†} and Michael Green^{4,5}

Table 2. Guideline-Endorsed Recommendations for the Prevention of EBV Disease and PTLD [1, 6].

| | SOT ^{a1} | HCT ^{b2} |
|--------------------------------|---|--|
| Prophylaxis | | |
| Chemoprophylaxis—Antivirals | Not recommended (weak/moderate to prevent EBV infection) (strong/moderate to prevent EBV disease) | Not recommended (DII) |
| Immunoprophylaxis | | |
| Vaccines | Unavailable | |
| IVIG | Not recommended (weak/moderate) | Not recommended (DIII) |
| Anti-CD20 | Not recommended (strong/low) | Marginally recommended (CII) |
| VSTs | Not recommended | Marginally recommended (CII) |
| Preemptive therapy | | |
| Reduction of immunosuppression | Recommended (strong/moderate for liver) (weak/low for other organs) | Recommended when combined with anti-CD20 (AII) |
| Chemoprophylaxis—Antivirals | Not recommended (weak/low) | Not recommended (DIII) |
| Immunoprophylaxis | | |
| Anti-CD20 | Not recommended (weak/very low) | Recommended, alongside RIS whenever possible (AII) |
| VSTs | Not recommended (weak/low) | Marginally recommended (CII) |

Abbreviations: EBV, Epstein-Barr virus; HCT, hematopoietic cell transplanation; IVIG, intravenous immunoglobulin; PTLD, post-transplant lymphoproliferative disorder, SOT, solid organ transplantation; VSTs, virus-specific T cells.

^aGrading recommendations for SOT: (x/y); x = strength of recommendation; y = quality of evidence.

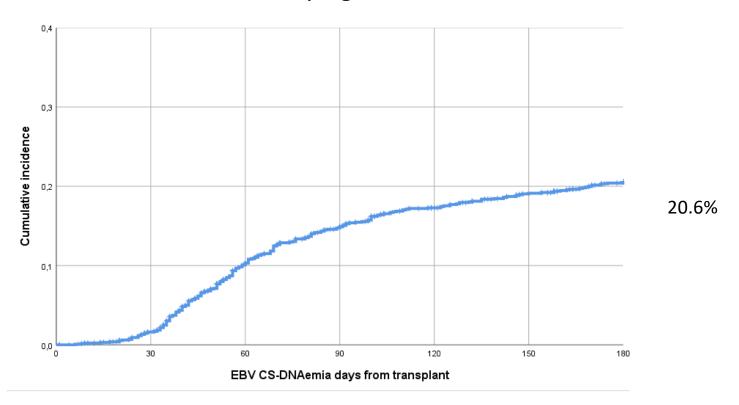
^bGrading recommendations for HCT: A = strong; B = moderate; C = marginal; D = against; I = at least 1 RCT; II = at least from one clinical trial; III = expert opinion, descriptive studies.



CLINICALTRIALS.GOV IDENTIFIER: NCT04412811



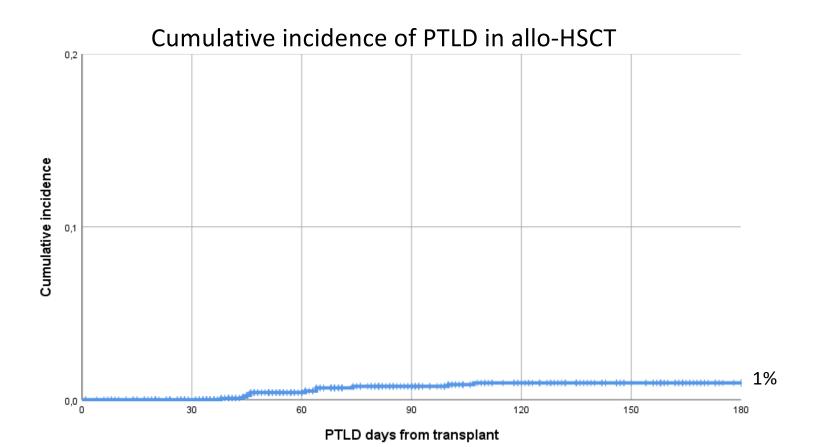
Cumulative incidence of clinically significant EBV DNAemia in allo-HSCT





CLINICALTRIALS.GOV IDENTIFIER: NCT04412811







CLINICALTRIALS.GOV IDENTIFIER: NCT04412811



Risk of CS-EBV DNAemia: variables considered in the analysis

- Age
- Sex
- Underlying disease: AL vs other
- Disease phase: CR, chronic, noCR
- Previous allo-HSCT
- Type of donor



- Conditioning regimen
- Stem cell source
- Letermovir prophylaxis

• T-cell depletion, ATG



- PT-CY
- R/D CMV serology
- ECOG PS
- HCT-CI
- Time of engraftment
- A-GVHD
- CS-CMV DNAemia

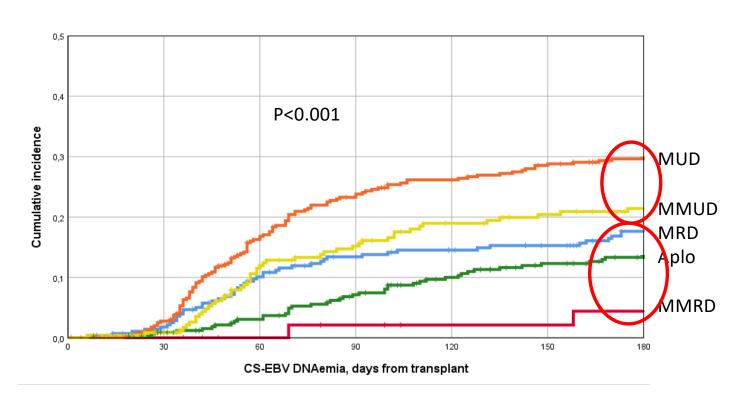




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Risk of CS-EBV DNAemia: type of transplant



PTLD: 11 cases

MUD: 7 cases

MMUD:2 cases

• MRD: 1 case

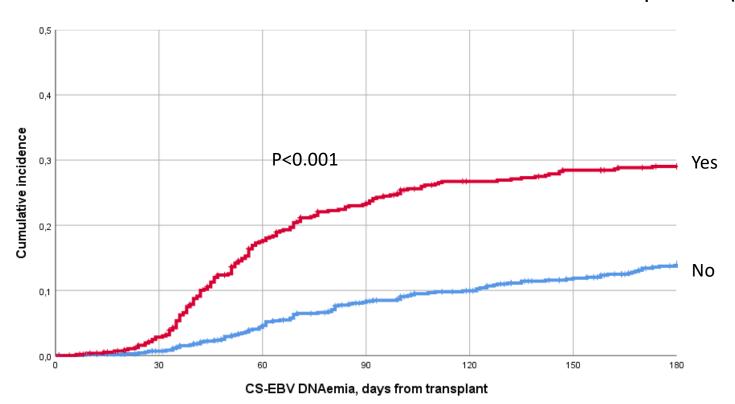
Aplo: 1 case



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Risk of CS-EBV DNAemia: T cell depletion (ATG)



PTLD: 11 cases

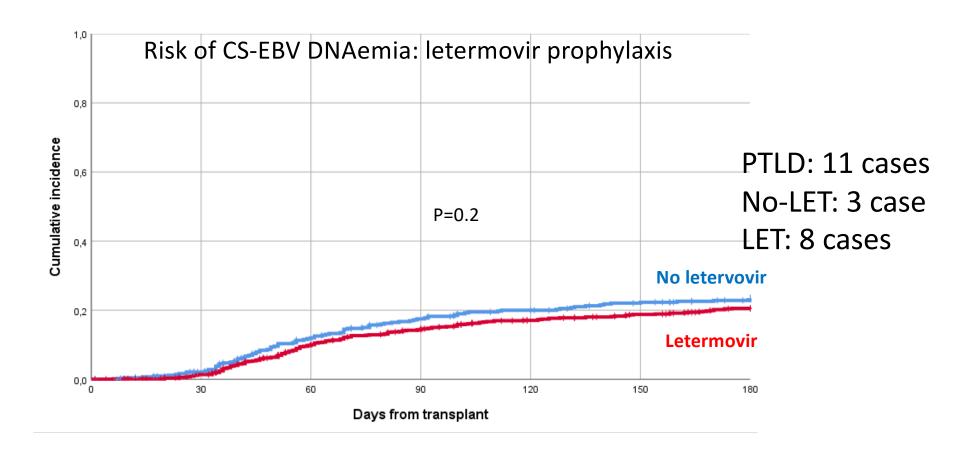
• TCD: 9 cases

No TCD:2 cases



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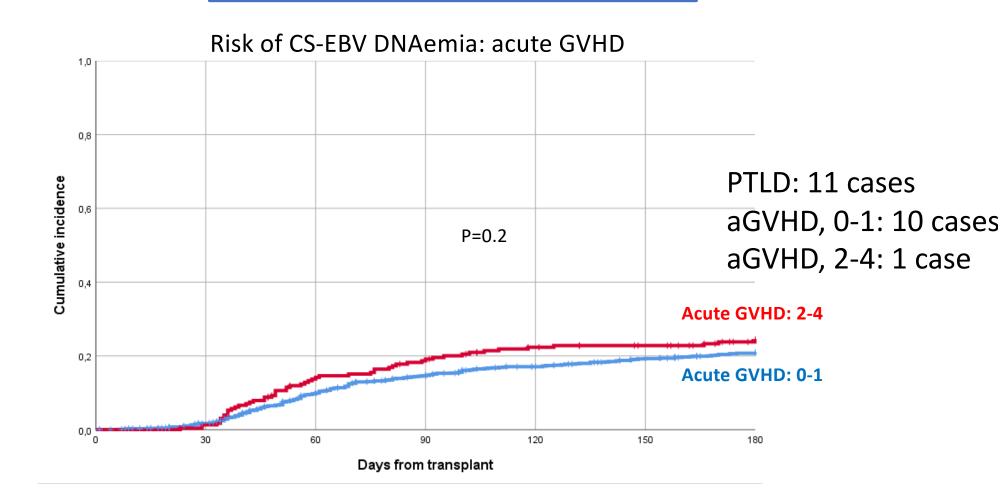






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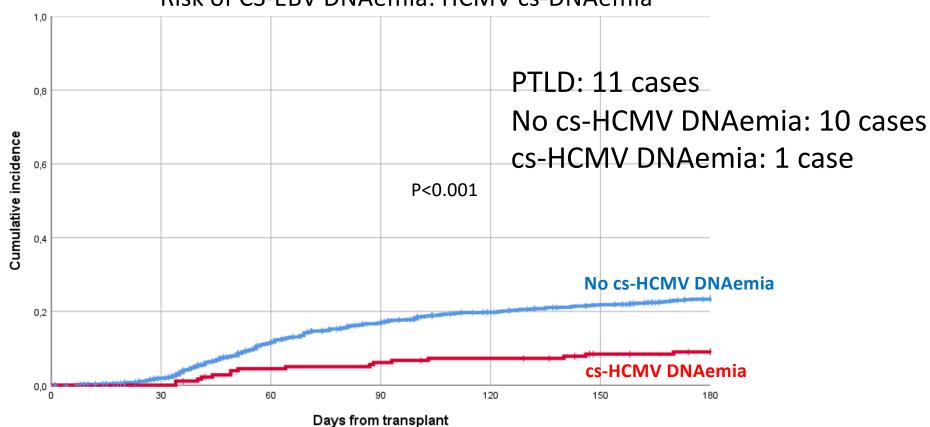












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Risk of CS-HCMV DNAemia, no letermovir

| Variables | | Univariate analysis | | Multivariate analysis | |
|-------------------------------------|---------------------|---------------------|---------|-----------------------|--------|
| | | HR (95% CI) | р | HR (95% CI) | р |
| Gender | Female | 1.00 | | | |
| | Male | 1.02 (0.63-1.66) | 0.93 | | |
| Age (increased by 10 years) | | 0.97 (0.96-0.98) | < 0.001 | | |
| 0 | <18 years | 1.00 | | | |
| Age | >= 18 years | 0.22 (0.13-0.35) | < 0.001 | 0.25 (0.15-0.42) | <0.001 |
| Underlying | Diseases other than | 1.00 | | | |
| hematologic | acute leukemia | | | | |
| disease | Acute leukemia | 0.67 (0.42-1.06) | 0.085 | | |
| Phase of the | Complete remission | 1.00 | | | |
| underlying disease | Chronic phase | 1.50 (0.83-2.69) | 0.18 | | |
| • • | No complete | 2.14 (1.22-3.77) | 0.008 | | |
| at transplant | remission | | | | |
| Previous HSCT | No | 1.00 | | | |
| | Previous auto-HSCT | 0.76 (0.25-2.33) | 0.64 | | |
| | Previous allo-HSCT | 1.74 (0.75-4.06) | 0.20 | | |
| CS-HCMV Infection | No | 1.00 | | | |
| in the 3 months | | 3.68 (1.00-13.6) | 0.051 | | |
| before transplant | Yes | | | | |
| | Negative/negative | 1.00 | | | |
| Recipient/donor | Negative/positive | 7.82 (0.97-62.8) | 0.053 | 7.80 (1.00-61.1) | 0.050 |
| HCMV serology | Positive/Negative | 33.1 (4.45-246) | < 0.001 | 31.5(4.31-229) | <0.001 |
| | Positive/Positive | 34.1 (4.75-245) | < 0.001 | 21.9 (3.06-157) | 0.002 |
| ECOG performance | 0-1 | 1.00 | | | |
| status at transplant | >1 | 0.66 (0.21-2.05) | 0.47 | | |
| HCT comorbidity index at transplant | Score 0 | 1.00 | | | |
| | Score 1-2 | 0.65 (0.36-1.18) | 0.16 | | |
| | Score >=3 | 0.39 (0.17-0.89) | 0.026 | | |
| Stem Cell Source | Peripheral blood | 1.00 | | | |
| | Bone marrow | 1.68 (1.04-2.71) | 0.033 | | |
| | Cord blood | 0.96 (0.15-6.20) | 0.97 | | |

| Variables | | Univariate | analysis | Multivariate analysis | |
|-----------------------------|---|-------------------|----------|-----------------------|-------|
| | | HR (95% CI) | р | HR (95% CI) | р |
| Donor type | Matched related | 1.00 | | | |
| | Mismatched related | 1.86 (0.71-4.90) | 0.21 | | |
| | Haploidentical | 1.59 (0.84-3.01) | 0.15 | | |
| | Matched unrelated | 1.07 (0.56- 2.06) | 0.84 | | |
| | Mismatched unrelated | 0.74 (0.31- 1.78) | 0.50 | | |
| | Myeloablative | 1.00 | | | |
| Conditioning regimen | Non myeloablative/ reduced intensity | 0.81 (0.47-1.40) | 0.46 | | |
| | No | 1.00 | | | |
| T cell depletion | Yes | 1.38 (0.87-2.21) | 0.18 | | |
| Use of post | No | 1.00 | | | |
| transplant | | 1.25 (0.75-2.09) | 0.40 | | |
| cyclophosphamide as | Yes | | | | |
| GVHD prophylaxis | | | | | |
| Days to engraftment | <=20 days | 1.00 | | | |
| Days to engraturient | >20 days | 1.11 (0.68-1.84) | 0.67 | | |
| Prophylaxis with CMV | No | 1.00 | | | |
| specific immunoglobulins | Yes | 0.47 (0.12-1.91) | 0.29 | | |
| Acute GvHD * | Grade 0-I | 1.00 | | | |
| | Grade II-IV | 1.43 (0.81-2.52) | 0.22 | | |
| EBV DNAemia* | Negative or <1000 copies/ml | 1.00 | | | |
| | >=1000 copies /ml | 0.33 (0.15-0.75) | 0.009 | 0.27 (0.11-0.62) | 0.002 |
| Gram negative * | No | 1.00 | | | |
| bacteremia | Yes | 0.81 (0.41-1.61) | 0.55 | | |
| Invasive fungal | No | 1.00 | | | |
| disease * | Yes | 0.34 (0.52-3.49) | 0.55 | | |

^{*} only cases observed before CS-HCMV infection were considered



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Has EBV a protective role against CS-HCMV-i in patients who do not receive LET-PP?

An immunological response to EBV infection may somehow interfere with HCMV reactivation.

- EBV is a polyclonal stimulator
- EBV levels and B-cell reconstitution were prospectively monitored in a cohort of allo-HSCT patients (Burns, Blood 2015). In patients with low or undetectable levels of EBV, the circulating B-cell pool consisted predominantly of transitional and naive cells, with a marked deficiency of CD27+memory cells. On the contrary, among patients with high EBV loads, there was a significant increase in both the proportion and number of CD27+ memory B cells.
- Some murine model studies showed that memory B cells can mediate protection against CMV in the absence of T cell help and transfer of memory B cells may be effective in protecting from an already ongoing viral infection (Winkler, Blood 2006; Klenovsek 2007).